



ESCOLA DE
MEDICINA

SCIENTIA MEDICA

Scientia Medica Porto Alegre, v. 35, p. 1-9, jan.-dez. 2025
e-ISSN: 1980-6108 | ISSN-L: 1806-5562

<http://dx.doi.org/10.15448/1980-6108.2025.1.46366>

SEÇÃO: ARTIGO ORIGINAL

Effect of the SARS-CoV-2 viral load on hematological and biochemical parameters

Efeitos da carga viral do SARS-CoV-2 nos parâmetros hematológicos e bioquímicos

Gustavo Caires Neves
Magalhães¹

orcid.org/0000-0001-7006-6732
cairesgustavo@yahoo.com.br

Clayson Moura Gomes²

orcid.org/0000-0001-8827-8274
claysonmoura10@gmail.com

Leandro do Prado
Assunção¹

orcid.org/0000-0002-1743-8151
leandrodoprado@discente.ufg.br

Sérgio Henrique
Nascente Costa¹

orcid.org/0000-0002-4225-6368
sergionascente@ufg.br

Fernando Antônio
Vinhais dos Santos³

orcid.org/0000-0009-0008-9047-9700
fernandovinhais@hlagyn.com

Keila Correia de
Alcântara¹

orcid.org/0002-4477-2833
keilalcantara@ufg.br

Received on: Jun 23, 2024.

Approved on: Nov 18, 2024.

Published on: 31 jan. 2025.



Artigo está licenciado sob forma de uma licença
Creative Commons Atribuição 4.0 Internacional.

Abstract

Objective: the viral load is determined by the cycle threshold (Ct), which is inversely proportional to the amount of target sequences present in the sample. In various viral diseases, the induction of inflammatory cytokines is strongly correlated with the viral load. However, the viral kinetics of SARS-CoV-2 remain poorly characterized, and its association with disease progression remains controversial. This study aimed to understand the correlation between the SARS-CoV-2 viral load and the biochemical and hematological markers in COVID-19-positive patients.

Methods: data from patients with detectable viral load for SARS-CoV-2 treated between March 2020 and May 2021 were collected between Jan/2023 and Jan/2024. Data were analyzed using Qui-square test, Student's t-test, and analysis of variance.

Results: the mean Ct values for the N and ORF1AB genes were 26.73 (± 3.95) and 25.93 (± 4.75), respectively. A Ct value below the mean for the N and ORF1AB genes, suggesting a higher viral load, was observed in 17.8% (n=152) and 18.4% (n=157) of the patients. The monocyte-to-lymphocyte ratio (MLR) was below the reference value for COVID-19-negative patients (P<0.0001). The C-reactive protein and ferritin levels were higher in patients with COVID-19 (P<0.05). Leucopenia (P<0.001), lymphocytopenia (P<0.001), neutropenia (P<0.001), and thrombocytopenia (P=0.003) were observed more frequently in patients with COVID-19.

Conclusion: the MLR below the reference value was more frequent observed in patients with a higher Ct value (lower viral load).

Keywords: COVID-19, RT-qPCR, biomarkers.

Resumo

Objetivo: a carga viral é determinada pelo *cycle threshold* (Ct), que é inversamente proporcional à quantidade de sequências-alvo presentes na amostra. Em várias doenças virais, a indução de citocinas inflamatórias está fortemente correlacionada com a carga viral. No entanto, a cinética viral do SARS-CoV-2 permanece pouco caracterizada e sua associação com a progressão da doença ainda é controversa. Este estudo teve como objetivo entender a correlação entre a carga viral do SARS-CoV-2 e os marcadores bioquímicos e hematológicos em pacientes positivos para COVID-19.

Métodos: os dados de pacientes com carga viral detectável para SARS-CoV-2 atendidos entre março de 2020 e maio de 2021 foram coletados entre janeiro de 2023 e janeiro de 2024. Os dados foram analisados utilizando o teste qui-quadrado, o teste t de Student e a análise de variância.

Resultados: os valores médios de Ct para os genes N e ORF1AB foram 26,73 ($\pm 3,95$) e 25,93 ($\pm 4,75$), respectivamente. Um valor de Ct abaixo da média para os genes N e ORF1AB, sugerindo uma carga viral mais alta, foi observado em 17,8% (n=152) e 18,4% (n=157) dos pacientes. A razão monócito-linfócito (MLR) estava abaixo do valor de referência para pacientes negativos para COVID-19 (P<0,0001). Os níveis de proteína C-reativa e ferritina encontravam-se mais altos em pacientes posi-

¹ Federal University of Goiás, Goiânia, GO, Brazil.

² Pontifical Catholic University of Goiás, Goiânia, GO, Brazil.

³ Transplant Immunology Laboratory – Hlagyn, Aparecida of Goiânia, GO, Brazil.

tivos para COVID-19 ($P < 0,05$). Leucopenia ($P < 0,0001$), linfocitopenia ($P < 0,0001$), neutropenia ($P < 0,0001$) e trombocitopenia ($P = 0,003$) foram observadas com mais frequência em pacientes positivos para COVID-19.

Conclusão: uma MLR abaixo do valor de referência foi observada com mais frequência em pacientes com um valor de Ct mais alto (carga viral mais baixa).

Palavras-chave: COVID-19, RT-qPCR, biomarcadores.

Introduction

Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) is the causative agent of COVID-19 and is responsible for more than six million deaths worldwide (1). In the acute phase of the disease, alterations in the hematological and inflammatory profiles occur, which is characterized by an increase in the levels of C-reactive protein (CRP), erythrocyte sedimentation rate, alanine aminotransferase, aspartate aminotransferase, lactate dehydrogenase, ferritin, and D-dimer, as well as evident lymphopenia (2). Biomarkers related to COVID-19 tend to gradually increase as the clinical condition worsens (3).

The association of disease progression with the viral kinetics of SARS-CoV-2 can aid in identifying high-risk patients and improving treatment strategies (4). Viral kinetics can be observed through reverse transcription quantitative polymerase chain reaction (RT-qPCR), where the viral load is established in the exponential phase of the reaction and is determined by the cycle threshold (Ct) (5). Ct represents the point at which the emitted fluorescence corresponds to the initial RNA quantity in the sample analyzed (6). Thus, the Ct value is inversely proportional to the number of target sequences present in the sample (viral load); in other words, the more copies, the lower the Ct value (7).

There is some correlation between the viral load and the clinical evolution of the disease in hospitalized patients with COVID-19; the viral load is higher in severe cases than in non-severe cases (8). Conversely, a study conducted in New York City determined that a higher viral load at the time of hospital admission was independently associated with mortality (9).

This study aimed to understand the association between the SARS-CoV-2 viral load and the hematological and biochemical markers in COVID-19-positive patients.

Materials and methods

A retrospective study was conducted using data from the clinical laboratory database of the Military Police Hospital of the State of Goiás. The requirement for obtaining informed consent was waived due to retrospective nature of the study and it was used deidentified data. The study was approved by the National Research Ethics Commission (No: 4.272.030).

The study included data collected from March 2020 to May 2021, a period of exponential increase in the number of COVID-19 cases in Brazil (10). Of the 12,648 individuals tested for SARS-CoV-2 using RT-qPCR at the Military Police Hospital of the State of Goiás, 853 were included in the study because they had undergone tests to determine complete blood count and CRP, ferritin, and D-dimer levels on the same date.

Ferritin, CRP and D-dimer were realized by Immunoturbidimetry (Mindray North America, Mahwah, NJ, USA). Reference values (RV) for ferritin were: children, 7.0–140.0 $\mu\text{g/L}$; men, 30.0–220.0 $\mu\text{g/L}$; and women, 20.0–110.0 $\mu\text{g/L}$, for CRP ≤ 8.5 mg/L and for D-dimer ≤ 600.0 ng/mL. All of them were categorized as normal or abnormal based on the RV.

Leukocyte and platelet counts were determined by impedance. The leukocytes were differentiated by optical flow cytometry and counted using a hematological counter (Celltac/MEK-7300; Nihon Kohden do Brasil). RV: Total leukocytes, 3,500–11,000/ μL ; lymphocytes, 800–4,000/ μL ; monocytes: 100–1,000/ μL ; neutrophils: 1,500–7,000/ μL ; platelets: 150,000–450,000/ μL . Absolute counts were categorized as normal, if within the RV. The abnormal values were further subdivided into high ($>RV$) and low ($<RV$). The neutrophil-to-lymphocyte ratio (NLR), platelet-to-lymphocyte ratio (PLR), and MLR were calculated.

RNA extraction and RT-qPCR were performed for the ORF1ab and N target genes using the XG-CV19-MB-g6-Mobius kit (Pinhais, Paraná, Brazil). Ct value <40 was considered RT-PCR/COVID-19 positive. Ct values were classified as "high viral load" when below the mean Ct value and "low viral load" when above the mean Ct value.

Data were tabulated and analyzed using Graph Prism (version 7 by Dotmatics), quantitative variables were analyzed using measures of central tendency. Qualitative variables were presented as absolute numbers and percentages. The chi-square test was used to verify if there was a significant association between categorical variables and analysis of variance (ANOVA) was performed to compare Ct values according to the hematological and biochemical parameters. A P-value <0.05 was considered statistically significant. Hypothesis testing was conducted using R Studio (version 4.2.1; Posit, MA, Boston for USA).

Among the 853 patients studied, a significant portion were male, and the majority fell within the age range of 18 to 65 years. The prevalence of SARS-CoV-2 infection was 33.8%, with males comprising more than half of the positive cases. In an analysis of COVID-19 positive patients, significant increases in CRP and ferritin levels were observed, exceeding the reference values in 90 (33.7%) and 183 (63.5%) cases, respectively. Furthermore, the frequency of hematological alterations was higher compared to patients without the infection. Leukopenia, lymphocytopenia, neutropenia, and thrombocytopenia were more prevalent among COVID-19 patients, indicating a significant impact of the disease on the hematological health of these individuals (**Table 1**). Approximately 89 (10.4%) of the included patients had an MLR less than the RV. Among the 89 patients, 74 (13.1%) tested negative for COVID-19 and 15 (5.2%) tested positive for COVID-19 (P = 0.0004) (Table 1).

Results

TABLE 1 – Biochemical and hematological markers in patients who tested positive and negative for COVID-19.

	Positive, n(%)	Negative, n(%)	Total, n(%)	P-value*
Total	288 (33.8)	565 (66.2)	853 (100)	
Sex female	132 (45.8)	272 (48.1)	404 (47.4)	0.52
Age, years				0.11
<18	10 (3.47)	18 (3.2)	28(3.3)	
18–65	254 (88.2)	520 (92.0)	774(90.7)	
>65	24 (8.3)	27 (4.8)	51(6)	
CPR >8.5 mg/L	90 (33.7)	132 (23.3)	222 (26.0)	0.01
D-dimer ≤600 ng/mL	228 (79.16)	451 (79.8)	679 (79.6)	0.82
Ferritin abnormal ^a	183 (63.5)	273 (48.3)	456 (53.5)	0.000
Leucocyte count, /μL				0.000
3,500–11,000	255 (88.0)	528 (93.0)	783 (91.8)	
>11,000	5 (1.7)	30 (5.3)	35 (4.1)	
<3,500	28 (9.7)	7 (1.2)	40 (4.1)	
Lymphocyte count, /μL				0.000
800–4,000	237 (82.2)	540 (95.6)	777 (91.1)	
>4,000	4 (1.4)	11 (1.9)	15 (1.8)	

TABLE 1 – Biochemical and hematological markers in patients who tested positive and negative for COVID-19. (CONT.).

	Positive, n(%)	Negative, n(%)	Total, n(%)	P-value*
MLR [†]				0.000
<800	47 (16.3)	14 (2.5)	61 (7.1)	
<0.125	15 (5.2)	74 (13.1)	89 (10.4)	
0.125–2.5	273 (94.8)	491 (86.9)	764 (89.6)	
Monocyte count, / μ L				0.38
100–1000	284 (98.6)	559 (98.9)	843 (98.8)	
>1000	3 (1.04)	6 (1.0)	9 (1.1)	
<100	1 (0.35)	0	1 (0.1)	
Neutrophil count, / μ L				0.000
1,500–7,000	262 (90.1)	510 (86.7)	772 (90.5)	
>7,000	6 (2.1)	49 (89.1)	55 (6.4)	
<1,500	20 (76.9)	6 (1.0)	26 (3.1)	
NLR				0.02
1.75–1.875	10 (3.5)	39 (6.9)	49 (5.7)	
>1.875	158 (54.9)	262 (46.4)	420 (49.2)	
<1.75	120 (41.6)	264 (46.7)	384 (45.1)	
Platelet count, / μ L				0.000
150,000–450,000	258 (89.6)	544 (96.3)	802 (94.0)	
>450,000	1(0.35)	4 (0.7)	5 (0.6)	
<150,000	27 (9.4)	17 (3.0)	44 (5.2)	
PLR				0.000
100.0–187.5	160 (55.5)	304 (53.8)	464 (54.4)	
>187.5	78 (27.1)	72 (12.7)	150 (17.6)	
<100	50 (17.4)	189 (33.4)	239 (28)	

CRP, C-reactive protein; NLR, neutrophil-to-lymphocyte ratio; PLR, platelet-to-lymphocyte ratio; MLR, mono-cyte-to-lymphocyte ratio.

*chi-square and[†] Fisher test. †No patient presented with MLR >2.5.

The average Ct values for the N and ORF1AB genes were 26.7 \pm 3.9 and 25.9 \pm 4.8, respectively. A Ct value below the average value for the N and ORF1AB genes, which suggests a higher viral load, was observed in 152 (17.8%) and 157 (18.4%) samples, respectively. When comparing

the Ct values between the group with abnormal results for inflammatory markers and the group with normal results, it was found that patients with an MLR below the RV had a lower viral load (a higher average Ct value) for both genes (P < 0.05) (**Table 2**).

TABLE 2 – Comparative analysis of the Ct values according to the hematological and biochemical parameters (n = 288).

	CT_N	P-value	CT_ORF1AB	P-value*
CPR		0.53		0.37
Normal	25.2±7.4		24.3±7.7	
Abnormal	25.7±6.3		25.1±6.6	
D-Dimer		0.23		0.26
Normal	25.6±7.0		24.9±7.3	
Abnormal	24.3±7.5		23.6±7.7	
Ferritin		0.69		0.79
Normal	25.2±7.2		24.5±7.5	
Abnormal	25.5±6.9		24.7±7.2	
Leucocytes		0.12		0.18
Normal	25.4±7.0		24.6±7.4	
Below	23.9±7.6		23.2±7.9	
Above	30.9±3.0		29.8±3.4	
Lymphocytes		0.78		0.91
Normal	25.4±7.3		24.6±7.6	
Below	24.9±6.3		24.2±6.6	
Above	27.1±4.3		25.6±4.7	
MLR		0.001		0.003
Normal	25.1±7.1		24.3±7.4	
Below	30.0±4.2		29.4±5.2	
Above	-		-	
Monocytes		0.62		0.67
Normal	25.3±7.1		24.5±7.4	
Below	30.1		26.7±4.1	
Above	28.015±3.8		30.1	
Neutrophils		0.09		0.08
Normal	25.4±7.2		24.6±7.5	
Below	23.5±6.2		22.2±6.3	
Above	30.7±3.7		29.8±4.4	
NLR		0.61		0.73
Normal	25.1±6.9		24.3±7.2	
Below	25.8±7.1		25.0±7.5	
Above	24.2±9.0		24.0±9.4	

TABLE 2 – Comparative analysis of the Ct values according to the hematological and biochemical parameters (n = 288). (CONT.).

	CT_N	P-value	CT_ORF1AB	P-value*
Platelets		0.35		0.40
Normal	25.5±7.0		23.7±7.3	
Below	24.0±7.8		23.3±8.2	
Above	31.6		29.8	
PLR		0.42		0.55
Normal	25.5±7.1		24.8±7.5	
Below	24.5±6.7		23.8±7.0	
Above	26.1±7.6		25.2±7.9	

CPR, C-Reactive Protein; MLR, monocyte-to-lymphocyte ratio; NLR, neutrophil-to-lymphocyte ratio; PLR, platelet-to-lymphocyte ratio. * ANOVA.

Discussion

RT-qPCR using respiratory samples is recommended by the WHO for diagnosing COVID-19 (12). Its semiquantitative measure, expressed by the Ct, indicates that a high Ct value represents a low viral load (13). Studies have explored the correlation of Ct values with disease severity and outcomes (14, 15). Ct is a potential prognostic marker (8) and is associated with clinical findings and laboratory parameters (16). However, the relationship between Ct values and the hematological and biochemical markers remains controversial. In our study, there was a significant change in the inflammatory marker levels in the patients with COVID-19, which is like the findings of other studies (17, 18) and it was found that the MLR was affected by the Ct value.

Studies have used the Ct value as an indirect measure of viral load (19, 20). A study in the U.S. determined that 676/4,428 (15.3%) of the samples that tested positive for COVID-19 had Ct values below the average value, suggesting a high viral load (21). In our study, Ct values below the average for N and ORF1AB genes were observed in 152 (17.8%) and 157 (18.4%) of the samples, respectively. However, although using Ct values obtained in molecular reactions as indirect measures of SARS-CoV-2 viral loads is simple, it may have errors, such as the cell or the sampled mucosal surface

mass not being considered (22).

The elevated levels of CRP and ferritin observed in a significant proportion of COVID-19 patients align with existing literature that identifies these markers as indicators of the excessive inflammatory response associated with viral infection (23). Both ferritin and CRP, as acute-phase proteins, are widely described as predictors of systemic inflammation and are linked to disease severity and prognosis in conditions such as viral pneumonia (23, 24). This excessive inflammatory pattern, evidenced by the higher prevalence of leukopenia, lymphocytopenia, and thrombocytopenia among COVID-19-positive patients, highlights a distinctive feature of the clinical profile of COVID-19, setting it apart from other respiratory infections (23). Thus, the use of these biomarkers may be valuable not only for diagnosis but also for risk stratification and clinical management of COVID-19 patients.

In the present study, we did not find a significant difference in the D-dimer levels between the COVID-19-positive and COVID-19-negative patients. This finding may be directly related to the fact that the patients included in this study were not hospitalized and did not demonstrate severe symptoms. An increase in this marker level has been associated with increased morbidity

and mortality (24, 25).

The NLR, MLR, and PLR are reportedly significantly higher among patients with COVID-19 than among normal individuals, especially in very severe cases. Thus, they are important prognostic markers for COVID-19 (26, 27). In our study, more COVID-19-positive patients had higher NLR and PLR values than COVID-19-negative patients. However, the MLR was within the normal range for many COVID-19-positive patients and below the RV for a significantly large number of COVID-19-negative patients.

In our study, a lower MLR was associated with a higher Ct value, indicating that in situations of lower viral load (higher Ct value), the immune response may be less intense (28, 29), which attenuates lymphocytopenia and decreases the MLR.

This was a retrospective study using secondary data from an outpatient clinic that lacked clinical data for a better evaluation of the studied population. Therefore, clinical outcomes could not be determined. Furthermore, the Ct value was assumed to be a direct measure of viral load because there are no standardized kits on the market for converting the Ct value to a normalized quantitative measure. Future studies evaluating the immune response according to viral load may demonstrate the correlation between the "cytokine storm" and lymphocytopenia, which consequently leads to an early increase in MLR. Thus, this ratio may be a prognostic marker in COVID-19.

Notes

This study is part of the result of a dissertation in the Postgraduate Program in Healthcare and Evaluation in Health from the Federal University of Goiás, by one of the authors (GCNG), in January, 2024, entitled "COVID-19: analyses of the influence of SARS-CoV-2 viral load on hematological and biochemical parameters".

Funding

This study did not receive financial support from external sources

Conflicts of interest disclosure

The authors declare no competing interests relevant to the content of this study.

Authors' contributions

All the authors declare to have made substantial contributions to the conception, or design, or acquisition, or analysis, or interpretation of data; and drafting the work or revising it critically for important intellectual content; and to approve the version to be published.

Availability of data and responsibility for the results

All the authors declare to have had full access to the available data and they assume full responsibility for the integrity of these results.

References

1. World Health Organization. WHO Coronavirus (COVID-19) dashboard [Internet]. 2020 [cited 2023 Jan 23]. Available at: <https://covid19.who.int>
2. Christensen B, Favaloro EJ, Lippi G, Van Cott EM. Hematology laboratory abnormalities in patients with coronavirus disease 2019 (COVID-19). *Semin Thromb Hemost.* 2020;46(07):845-9. <https://doi.org/10.1055/s-0040-1715458>
3. Lippi G, Plebani M. Laboratory abnormalities in patients with COVID-2019 infection. *Clin Chem Lab Med (CCLM).* 2020;58(7):1131-4. <https://doi.org/10.1515/cclm-2020-0198>
4. Contreras C, Newby JM, Hillen T. Personalized virus load curves for acute viral infections. *Viruses.* 2021;13(9):1815. <https://doi.org/10.3390/v13091815>
5. Ladeira PR, Isaac C, Ferreira MC. Reação em cadeia da polimerase da transcrição reversa em tempo real. *Rev. Med.* 2011;90(1):47. <https://doi.org/10.11606/issn.1679-9836.v90i1p47-51>
6. Wong ML, Medrano JF. Real-time PCR for mRNA quantitation. *BioTechniques.* 2005;39(1):75-85. <https://doi.org/10.2144/05391rv01>
7. Menezes ME, Lima LM, Martinello F. Diagnóstico laboratorial do SARS-CoV-2 por transcrição reversa seguida de reação em cadeia da polimerase em tempo real (RT-PCR). *Rev Bras Anal Clin.* 2020;52(2):122-30. <https://doi.org/10.21877/2448-3877.20200006>

8. Liu Y, Yan LM, Wan L, Xiang TX, Le A, Liu JM, et al. Viral dynamics in mild and severe cases of COVID-19. *Lancet Infect Dis.* 2020;20(6):656-7. [https://doi.org/10.1016/S1473-3099\(20\)30232-2](https://doi.org/10.1016/S1473-3099(20)30232-2)
9. Magleby R, Westblade LF, Trzebucki A, Simon MS, Rajan M, Park J, et al. Impact of severe acute respiratory syndrome coronavirus 2 viral load on risk of intubation and mortality among hospitalized patients with coronavirus disease 2019. *Clin Infect Dis.* 2020;73(11):e4197-205. <https://doi.org/10.1093/cid/ciaa851>
10. Brasil. Ministério da Saúde. Paineis do Coronavírus [Internet]. 2023 [cited 2023 May 09]. Available at: <https://covid.saude.gov.br>
11. Azzi L, Carcano G, Gianfagna F, Grossi P, Gasperina DD, Genoni A, et al. Saliva is a reliable tool to detect SARS-CoV-2. *J Infect.* 2020;81(1):e45-e50. <https://doi.org/10.1016/j.jinf.2020.04.005>
12. World Health Organization. Laboratory testing strategy recommendations for COVID-19 [Internet]. Geneva: World Health Organization; 2020 [cited 2023 Nov 21]. Available at: <https://apps.who.int/iris/handle/10665/331509>
13. Walsh KA, Jordan K, Clyne B, Rohde D, Drummond L, Byrne P, et al. SARS-CoV-2 detection, viral load and infectivity over the course of an infection. *J Infect.* 2020;81(3):357-71. <https://doi.org/10.1016/j.jinf.2020.06.067>
14. Rabaan AA, Tirupathi R, Sule AA, Aldali J, Mutair AA, Alhumaid S, et al. Viral dynamics and Real-Time RT-PCR Ct values correlation with disease severity in COVID-19. *Diagnostics.* 2021;11(6):1091. <https://doi.org/10.3390/diagnostics11061091>
15. Rao SN, Manissero D, Steele VR, Pareja J. A narrative systematic review of the clinical utility of cycle threshold values in the context of COVID-19. *Infect Dis Ther.* 2020;9(3):573-86. <https://doi.org/10.1007/s40121-020-00324-3>
16. Ataee Z, Rahmani A, Amel S, Khadem-Rezaian M, Ziaee M. Relationship of viral load with the laboratory markers and prognosis in COVID-19 patients. *Med J Islam Repub Iran.* 2023;37(1):544-8. <https://doi.org/10.47176/mjiri.37.67>
17. Elshazli RM, Toraih EA, Elgaml A, El-Mowafy M, El-Mesery M, Amin MN, et al. Diagnostic and prognostic value of hematological and immunological markers in COVID-19 infection: a meta-analysis of 6320 patients. *PLoS ONE.* 2020;15(8):e0238160. <https://doi.org/10.1371/journal.pone.0238160>
18. Huang I, Pranata R, Lim MA, Oehadian A, Alisjahbana B. C-reactive protein, procalcitonin, D-dimer, and ferritin in severe coronavirus disease-2019: a meta-analysis. *Ther Adv Respir Dis.* 2020;14:1-14. <https://doi.org/10.1177/1753466620937175>
19. Asai N, Sakanashi D, Ohashi W, Nakamura A, Yamada A, Kawamoto Y, et al. Could threshold cycle value correctly reflect the severity of novel coronavirus disease 2019 (COVID-19)? *J Infect Chemother.* 2021;27(1):117-9. <https://doi.org/10.1016/j.jiac.2020.09.010>
20. Shoaib N, Iqbal A, Shah FA, Zainab W, Qasim M, Zerqoon N, et al. Population-level median cycle threshold (Ct) values for asymptomatic COVID-19 cases can predict the trajectory of future cases. *PLoS ONE.* 2021;16(3):e0281899. <https://doi.org/10.1371/journal.pone.0281899>
21. Kleiboeker S, Cowden S, Grantham J, Nutt J, Tyler A, Berg A, et al. SARS-CoV-2 viral load assessment in respiratory samples. *J Clin Virol.* 2020;129:104439. <https://doi.org/10.1016/j.jcv.2020.104439>
22. Dahdouh E, Lázaro-Perona F, Romero-Gómez MP, Mingorance J, García-Rodríguez J. Ct values from SARS-CoV-2 diagnostic PCR assays should not be used as direct estimates of viral load. *J Infect.* 2021;82(3):414-51. <https://doi.org/10.1016/j.jinf.2020.10.017>
23. Parimoo A, Biswas A, Baitha U, Gupta G, Pandey S, Ranjan P, et al. Dynamics of inflammatory markers in predicting mortality in COVID-19. *Cureus.* 2021;13(10):e19080. <https://doi.org/10.7759/cureus.19080>
24. Chen T, Wu D, Chen H, Yan W, Yang D, Chen G, et al. Clinical characteristics of 113 deceased patients with coronavirus disease 2019: retrospective study. *BMJ.* 2020;368:m1091. <https://doi.org/10.1136/bmj.m1091>
25. Şener G, Bayrak T, Coşkun C, Bayrak A. Neutrophil lymphocyte ratio, monocyte lymphocyte ratio, platelet lymphocyte ratio in Covid-19 patients. *Clin Lab.* 2022;68. <https://doi.org/10.7754/clin.lab.2021.210639>
26. Bull S, Jamrozik E, Binik A, Parker MJ. SARS-CoV-2 challenge studies: ethics and risk minimisation. *J. Med. Ethics.* 2021;47:e79. <https://doi.org/10.1136/medethics-2020-106504>
27. Peng J, Qi D, Yuan G, Deng X, Mei Y, Feng L, et al. Diagnostic value of peripheral hematologic markers for coronavirus disease 2019 (COVID19): a multicenter, cross-sectional study. *J Clin Lab Anal.* 2020;34:e23475. <https://doi.org/10.1002/jcla.23475>
28. Channappanavar R, Perlman S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. *Semin Immunopathol.* 2017;39(5):529-39. <https://doi.org/10.1007/s00281-017-0629-x>
29. Felsenstein S, Herbert JA, McNamara PS, Hedrich CM. COVID-19: immunology and treatment options. *Clin Immunol.* 2020;215:108448. <https://doi.org/10.1016/j.clim.2020.108448>

Gustavo Caires Neves Magalhães

MSc in Healthcare and Evaluation in Health from the Postgraduate Program in Healthcare and Evaluation in Health from Federal University of Goiás (UFG), in Goiânia, GO, Brazil.

Clayson Moura Gomes

PhD in Tropical Medicine and Public Health from the Federal University of Goiás (UFG), in Goiânia, GO, Brazil. Professor at Pontifical Catholic University of Goiás (PUC-GO) in Goiânia, GO, Brazil.

Leandro do Prado Assunção

PhD in Tropical Medicine and Public Health from the Federal University of Goiás (UFG) in Goiânia, GO, Brazil.

Sérgio Henrique Nascente Costa

PhD in Health Science from Federal University of Goiás (UFG), in Goiânia, GO, Brazil. Professor at the Pharmacy Faculty from Federal University of Goiás (UFG), in Goiânia, GO, Brazil.

Fernando Antônio Vinhal dos Santos

PhD in Applicable Immunology and Parasitology from the Federal University of Uberlândia (UFU), in Uberlândia, MG, Brazil. Technical Director of the Transplant Immunology Laboratory – Hlagyn, in Aparecida of Goiânia, GO, Brazil.

Keila Correia de Alcântara

PhD in Tropical Medicine and Public Health from the Federal University of Goiás (UFG), in Goiânia, GO, Brazil. Professor at the Pharmacy Faculty from Federal University of Goiás (UFG), in Goiânia, GO, Brazil.

Mailing Address

Dra. Keila Correia de Alcântara

Universidade Federal de Goiás, Faculdade de Farmácia
Campus Universitário, 74605220
Goiânia, GO, Brasil